DifferInt

Compositional differentiation among populations at three levels of genetic integration

User Manual

May 2013

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Gillet, E.M. (2013) DifferInt: Compositional differentiation among populations at three levels of genetic integration. *Molecular Ecology Resources* 13, 953-964. http://dx.doi.org/10.1111/1755-0998.12145

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1 Introduction

The computer program *DifferInt* is described in the publication

Gillet, E.M. (2013) DifferInt: Compositional differentiation among populations at three levels of genetic integration. Molecular Ecology Resources 13, 953-964. http://dx.doi.org/10.1111/1755-0998. 12145

It calculates four measures of compositional differentiation among populations for their genetic types at three levels of genetic integration: the gene pool level, the mean single-locus level and the multilocus level. Genetic types refer to the single-locus or multilocus genotypes of diploid individuals in one or more populations or population samples at a given set of codominantly expressed marker loci.

1.1 Minimality principle

All four measures are based on the minimality principle, which states that the distance between two populations equals the minimum extent to which one population must be altered in order to make it match the other based on two tiers of dissimilarity:

- dissimilarity between two individuals as the proportion of genes in the genetic type of one individual that must be changed in order to make this individual match the other, termed the elementary genic difference d,
- dissimilarity between two populations as the minimum extent to which the frequencies of genetic types in one population must be altered in order to make this population match a second.

1.2 Pairwise genetic distance

Considering only the frequencies (p_1, \ldots, p_n) and (q_1, \ldots, q_n) of the genetic types in two populations \mathcal{P} and \mathcal{Q} at any given level, the absolute genetic

distance

$$d_0(\mathcal{P}, \mathcal{Q}) = \frac{1}{2} \sum_i |p_i - q_i|$$

measures the minimum proportion of genetic types in either one of the two populations that must be shifted to a different type in order to make this population match the other.

The minimality principle implies that genetic types showing a frequency excess in the one population should be shifted to frequency-deficient genetic types that differ in as few genes as possible, that is, minimize the elementary genic difference d. Simultaneous minimization of the extent of alteration with respect to both frequency differences and elementary genic differences yields the measure of pairwise population distance Δ . Its calculation requires application of a linear optimization algorithm. This approach and its implications for the measurement of gene associations is described in the open-access publication by Gillet & Gregorius (2008).

1.3 Genetic differentiation

The pairwise genetic distances Δ and d_0 are generalized to differentiation measures for higher numbers of populations in two ways:

- Complementary differentiation as the mean contribution of genetic types by each population to the genetic types represented in the entirety of all populations,
- Dispersive differentiation as the mean distance of the genetic types contributed by each population from the contributions of each of the others.

DifferInt calculates the following differentiation measures (see Tab. 1), all of which were developed within our working group:

• complementary genetic differentiation Δ_{SD} that considers elementary genic differences d, together with the contributions $\Delta_{SD}(j)$ of each population $\mathcal{P}(j)$,

	Pairwise distance	Complementary differentiation	Dispersive differentiation
Considering elementary genic differences	Δ	Δ_{SD}	$\bar{\Delta}$
Neglecting elementary genic differences	d_0	δ_{SD}	$\bar{\delta}$

Table 1: Measures of pairwise genetic distance between two populations and the two corresponding measures of genetic differentiation among populations. All measures are calculated by *DifferInt*/ at three levels of genetic integration: gene-pool, single-locus genotypes, and multilocus genotypes.

- a new measure of dispersive differentiation Δ that considers elementary genic differences d, together with the contributions $\overline{\Delta}(j)$ of each population $\mathcal{P}(j)$
- complementary differentiation δ_{SD} that neglects elementary genic differences, together with the contributions $\delta_{SD}(j)$ of the $\mathcal{P}(j)$ th population
- a new measure of dispersive differentiation $\bar{\delta}$ that neglects elementary genic differences, together with the contributions $\bar{\delta}_{SD}(j)$ of the $\mathcal{P}(j)$ th population.

1.4 Levels of genetic integration

In order to assess the effects of gene association, both homologous (*i.e.*, intralocus) and non-homologous (*i.e.*, interlocus), on genetic differentiation, all measures are calculated at three levels of genetic integration: the genepool, single-locus genotypes, and multilocus genotypes. A special property of the genetic distance Δ is that it cannot decrease from one level of integration to the next highest (see Gillet & Gregorius (2008) for mathematical proof; also see App. A). It follows that Δ_{SD} and $\overline{\Delta}$ also cannot decrease from one level to a higher level.

This property allows inference on the similarity of forms of association within pairs of populations, since it remains equal if the genotypes in the two populations are composed by the same form of gene association. $\overline{\Delta}$ also remains equal if the genotypes in the two populations are composed by the same form of gene association. This is not true of Δ_{SD} , since the mixture of populations with the same form of association adds new association to the complement of a population, e.g. the increase in homozygosity in mixtures of populations that show Hardy-Weinberg-Proportions (Wahlund-Effect).

1.5 Snail diagrams

Subpopulation differentiation is illustrated by so-called snail diagrams. Each population is represented by a sector of a pie-like chart. The radius of the sector equals the contribution of one of the populations to differentiation (*i.e.*, $\Delta_{SD}(j)$, $\bar{\Delta}(j)$, $\delta_{SD}(j)$, or $\bar{\delta}(j)$), while the angle of the sector is proportional to the relative size of this population among all populations. The sectors are arranged by radius, with the largest contribution placed to the right of 90° and the sector with the second-largest radius to its right, etc., yielding a figure that is reminiscent of the shell of a snail. A dotted circle of radius equal to the weighted mean of the sector radii marks the population differentiation (*i.e.*, Δ_{SD} , $\bar{\Delta}$, δ_{SD} , or $\bar{\delta}$). The snails are encoded as *fig*-files for vector graphic software *WinFIG* (http://www.winfig.com) for Windows or *xfig* (http://www.xfig.org) for Linux. Optionally, snails are displayed on the screen. Examples are shown in Figs. 4-7.

1.6 Covariation of differentiation

A measure of covariation C expresses the degree of correspondence between the contributions of the single populations to differentiation over two levels of integration (see Sec. 7.4).

1.7 Statistical analysis

Statistical significance is estimated using two kinds of permutation analysis (see Sec. 7.5):

• Permutation of alleles among individuals within populations tests the homologous association of alleles into single-locus genotypes as well as

the non-homologous association of single-locus genotypes into multilocus genotypes.

• Permutation of multilocus genotypes among populations tests the association, or partitioning, of genetic types among the populations.

Results are expressed as confidence intervals of chosen percentage of permutation values, minimum and maximum permutation values, and P-values.

2 Citing DifferInt

Gillet, E.M. (2013) DifferInt: Compositional differentiation of populations at three levels of genetic integration. *Molecular Ecology Resources* 13, 953-964. http://dx.doi.org/10.1111/1755-0998.12145

3 Installation

3.1 Installing *DifferInt*

Download the current version of *DifferInt* from either of the following websites:

```
http://www.uni-goettingen.de/forstgenetik/differint
https://projects.gwdg.de/projects/differint
```

• Windows (32-bit)

Download the file DifferInt-W32.zip to the desired directory and unzip it. A new directory named DifferInt-W32 is created in the current directory that contains the following files

```
DifferInt.exe
exa1.txt
exa2.txt
Gillet-Gregorius-BMCGenetics-2008.pdf
metapop1.dat
metapop2.dat
metapop3.dat
metapop4.dat
GNU-General-Public-License.txt
README
```

Program executable First example input file Second example input file Original publication Example M^A in Gillet (2013) Example M^B in Gillet (2013) Example M^C in Gillet (2013) Example M^D in Gillet (2013) GNU General Public License Instructions, notes and bug fixes

• Linux

Download DifferInt-linux.tar to the desired directory and unpack it, e.g. by opening terminal window in this directory and typing

> tar -xvf DifferInt-linux.tar

A new directory named DifferInt-linux is created in the current directory that contains the following files

DifferInt	Program executable
exa1.txt	First example input file
exa2.txt	Second example input file
Gillet-Gregorius-BMCGenetics-2008.pdf	Original publication
GNU-General-Public-License.txt	GNU General Public License
metapop1.dat	Example M^A in Gillet (2013)
metapop2.dat	Example M^B in Gillet (2013)
metapop3.dat	Example M^C in Gillet (2013)
metapop4.dat	Example M^D in Gillet (2013)
README	Instructions, notes and bug fixes

3.2 Installing plot software

If selected in the input menu, *DifferInt* produces snail diagrams (see Figs.4-7) as files with names ending in .fig. To display, print, and alter these diagrams, additional software must be installed on the computer (xxx stands for the current version number):

• For Windows, DifferInt displays snails by calling the shareware Win-FIG. Install WinFIG and WinFIG_fontspack_1.0.zip as described. • For Linux, *DifferInt* displays snails by calling the open-access software *xfig* and *transfig*. If they are not included in your Linux distribution, install them from http://www.xfig.org as described. Installation in the directory /usr/local/lib requires root authorization.

4 Input file

4.1 Construction of input file

A file containing the genetic data is prepared in advance using a text editor or spreadsheet software. Consecutive columns of data may be separated by any number of blanks, a single comma, or a single TAB. When finished, save or export the data as a text-only(!) file (i.e., without formatting) named, for example, input.txt. Place it in the same directory as the executable.

Line-by-line description of input files:

- * Line 1: At least one character "#" followed by name of data
- * Line 2: Number of populations
- * Line 3: Number of gene loci
- * Line 4: "yes", if each line in the list of genetic types is to begin with the count of individuals that possess this genetic type; "no" otherwise
- * Line 5: Empty line (optional)
- * Line 6: At least one character "#" followed by name of population 1 (optional)
- * Lines 7ff: Population 1
 - If line 4 contains "yes", each subsequent line contains a number of individuals (integer) of the same genetic type followed by specification of this (multilocus) genotype as a list of its two alleles at gene locus 1, then gene locus 2, etc. Alleles are designated by up to 3 numerical digits (integers). See example input file exal.txt in Fig. 1.

Figure 1: Example input file exa1.txt

```
#Example1
       #No. populations
3
2
       #No. loci
       #Each genotype is preceded by number of individuals (yes|no)
yes
#Population 1
       1 1
                1 1
2
2
                1 2
       1 1
                \frac{1}{2} \frac{1}{2}
3
3
2
3
3
2
2
2
       1 1
       1 2
1 2
                1 1
                1 2
       1 2
2 2
                2 2
                1 1
       2 2
3 3
                1 2
                2 3
#Population 2
2 1 1 1 1
               1 2
2 2
1
       1 1
1
       1 1
1
       1 2
                1 1
       1 2
1 2
2 2
2 2
2 2
2 2
                1 2
1
                2 2
1
                1 1
1
                1 2
1
                \bar{2} \bar{2}
1
#Population 3
       1 1
                1 1
2
2
2
2
2
2
2
2
2
2
2
       1 1
                33
                4 4
       1 1
       1 3
                1 1
       1 3
                1 2
               4 4
1 2
       1 4
2 2
       2 3
                2 3
2
       4 4
                4 4
```

Figure 2: Example input file <code>exa2.txt</code> – same data as in Fig. 1

#Example1 3 #No. 2 #No. no #Each	population loci genotype i	s preceded	by number	of ind	lividuals	(yes no)
<pre>#Populati 1 1 1 1 1 1 1 1 1 1 1 1 2 1 1 1 2 1 1 1 2 1 1 2 2 1 1 2 2 1 1 2 2 1 1 2 2 1 1 2 2 1 2 1 1 1 2 1 1 1 2 1 1 1 2 1 2 1 2 2 2 1 2 2 2 1 2 2 2 1 2 2 2 1 2 2 2 1 2 2 2 1 1 2 2 1 1 2 2 1 1 2 2 1 2 3 3 2 3 3 3 2 3</pre>	on 1			#Popu 1 1 1 1 1 1 1 1 1 1 1 1 1 1	11ation 3 1 1 1 1 3 3 3 3 4 4 4 4	
<pre>#Populati 1 1 1 1 1 1 1 1 1 1 1 2 1 1 2 2 1 2 1 2 1 2 1 2 1 2 2 2 2 2 1 1 2 2 1 2 2 2 2 1 2 2 2 2 2</pre>	on 2			$ \begin{array}{c} 1 & 3 \\ 1 & 3 \\ 1 & 3 \\ 1 & 3 \\ 1 & 4 \\ 2 & 2 \\ 2 & 3 \\ 4 & 4 \\ 4 & 4 \end{array} $	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	

[input file continued to the right] \nearrow

- If line 4 contains "no", each subsequent line contains only the (multilocus) genotype of a single individual, as specified in the previous paragraph. Thus two individuals possessing the same genetic type occupy two lines of input. See example input file exa2.txt in Fig. 2.
- * Empty line
- $\ast\,$ For each subsequent population, repeat previous lines beginning with line 6
- * End file with one empty line (or one line filled with any number of blanks)

4.2 Example of input files

The two example input files exa1.txt in Fig. 1 and exa2.txt in Fig. 2 are equivalent in that they show the same data. In exa1.txt, individuals possessing the same genotype are collected into one line headed by their number. In exa2.txt, each individual occupies a line of its own with its genotype.

5 Program execution

Execution of *DifferInt* is initiated by one of three methods:

5.1 Menu-driven execution

5.1.1 Start program

In a file manager, start execution by double-clicking on

```
DifferInt.exe (for Windows)
DifferInt (for Linux)
```

or in a terminal window, start execution by typing

> DifferInt (for Windows)

> ./DifferInt (for Linux)

!!!!!!!! IMPORTANT NOTE FOR LINUX USERS !!!!!!!!!

On some computers, warnings of the following type appear

```
Warning: translation table syntax error: Unknown keysym name:
osfActivate
Warning: ... found while parsing ':<Key>osfActivate:
ManagerParentActivate()'
```

As explained by Helmut Michels (2011) in the DISLIN Discussion Group, when a program that uses DISLIN widgets is executed on a recent or older Linux system than the one on which it was created, warnings of this type may appear. The reason is that the location of the file XKeysymDB used by the widgets can differ between Linux systems. A workaround is to define the environment variable XKEYSYMDB with the location of the XKeysymDB file. Thus if

> find /usr/ -name XKeysymDB

or

> locate XKeysymDB

returns location, then enter

> export XKEYSYMDB=location

For example, if location is /usr/share/X11/XKeysymDB, then type

> export XKEYSYMDB=/usr/share/X11/XKeysymDB

and restart the program by typing

> ./DifferInt

5.1.2 File select window

A window opens for selection of the input file, e.g. input.txt. Close window by clicking OK.

nput file: /home/egillet/fortran/Delta/DifferInt-2.0/src-DifferInt-2.0/linu:	e of
	2 a.
ist one or more loci for inclusion in multilocus genotypes	All 2 loci
ermute alleles over individuals within populations ?	🗄 Yes 🗇 No
Permute individual genotypes among populations ?	🛇 Yes 🛇 No
Number of permutations ?	
Size of confidence intervals as percentage of permutations	95.0
Prefix for names of output files (max.10 characters) ?	
Make snail graphics (using "xfig" software) ?	🔷 Yes 💠 No
ОК	

Figure 3: Interactive input window for menu-driven execution

5.1.3 Interactive input window

An interactive input window opens (see Fig. 3) requesting the following specifications:

- * List one or more loci for inclusion in multilocus genotypes: To select all loci, retain preselected text "All X loci". Otherwise, replace text by a list of loci by number, separated by blanks or commas. The number of loci in the data is determined by a preliminary reading of the input file.
- * Permute alleles over individuals within populations ?: Select "Yes" or "No" (see Sec.7.5.1).
- * Permute individual genotypes among populations ?: Select "Yes" or "No" (see Sec.7.5.2).
- * Number of permutations?:

Insert a large integer. There is no default value.

 st Size of confidence intervals as percentage of permutations:

Either retain preselected value of "95.0" or replace by desired percentage "X" (with or without decimal point).

The confidence interval is a result of permutation analysis. It is taken to span the central X% of the parameter values yielded by all permutations. Thus the $\frac{1}{2} \cdot X\%$ smallest and the $\frac{1}{2} \cdot X\%$ largest values are excluded from the confidence interval.

* Prefix for names of output files (max.10 characters)?:

Insert up to 10 characters to designate output files. If, for example, "aaa", is entered, the names of all output files will begin with "aaa". There is no default prefix.

* Make snail graphics (using WinFig software?) (for Windows) or

Make snail graphics (using xfig software)? (for Linux):

For each level of genetic integration, the snails rank the populations as pie pieces of decreasing radius around a central point, where the radius assigned to each population is proportional to the genetic distance of this population to the complement population formed by pooling all other populations (Gillet & Gregorius, 2008; Gregorius & Roberds, 1986).

If the answer is "yes", two files are prepared for the creation of snail graphics (for prefix "aaa") that serve as input to the vector graphics program *xfig* (for Linux) or *WinFig* (for Windows) (see Sec. 3.2).

aaa.Delta.schnecken.fig: These snails represent the genetic distances $\Delta_{SD}(j)$, which consider the elementary genic differences between genetic types

aaa.d0.schnecken.fig: These snails represent the genetic distances D_j , which neglect genic differences

If the answer is "no", the two .fig-files are produced but not displayed. They can be called up later and modified by one of the vector graphics programs *WinFIG* for Windows or *xfig* for Linux (see Sec. 3.2).

5.2 Keyboard-driven execution

This form of execution is initiated by starting DifferInt with the option -nomenu, *i.e.*,

> DifferInt -nomenu (for Windows)

> ./DifferInt -nomenu (for Linux)

A line then appears asking for the name of the input file:

> Input name of input file :

Enter name of file and press <Return>.

A listing of the program's heading appears, followed by a series of questions to be answered over the keyboard. The questions are the same as for menudriven execution (see Sec. 5.1.3).

5.3 Batch-type execution

A file (e.g. batch.txt) can be prepared in advance with a text editor that contains the answers to all of the questions posed under keyboard-driven execution, just as if they were being typed directly into the terminal window. DifferInt is started by typing into a terminal window (without a minus-sign)

> DifferInt batch.txt (for Windows)
> ./DifferInt batch.txt (for Linux)

6 Command-line arguments

When execution is initiated from a terminal window, up to three arguments can be added on the command line (brackets [] signify that inclusion is optional)

```
> DifferInt [arg1 [arg2 [arg3] ] ] (for Windows)
> ./DifferInt [arg1 [arg2 [arg3] ] ] (for Linux)
```

The arguments can be of the following forms:

- If arg1 does not begin with a minus sign, it is taken to be the name of a batch file containing all specifications (e.g. the file batch.txt discussed in Sec. 5.3). Additional command-line arguments may follow, but this must be the first.
- -nomenu

Name of input file and program specifications are to be requested through the keyboard (see Sec. 5.2).

• -ncols=X

The positive integer X specifies the maximum number of columns that are printed on a line in the output file. If the results contain more than X columns, the line is wrapped. By default, -ncols=10.

7 Calculations

DifferInt calculates the following measures at all three levels of genetic integration, then determines confidence intervals and P-values by two forms of permutation analysis.

7.1 Levels of genetic integration

Genetic integration signifies the form of association of genes to single-locus genotypes to multilocus genotypes. *DifferInt* considers three levels of genetic integration:

7.1.1 Gene-pool

The lowest level of genetic integration is the gene-pool. The gene-type of each individual gene is characterized by the gene locus at which it is located and by its allelic state. Assuming that the degree of ploidy is the same at all loci, the relative frequencies of the gene-types in the gene-pool of a population equal $(1/L) \cdot p_{i;l}$, where L is the number of loci L and $p_{i;l}$ is the relative frequency of the *i*-th allele at the *l*-th gene locus in the population $(\sum_i p_{i;l} = 1 \text{ and } \sum_{i,l} (1/L) \cdot p_{i;l} = 1).$

7.1.2 Single-locus genotypes

In polyploid species, more than one gene is present at the same gene locus. In diploid species, two genes are present at all (autosomal) loci. Thus the genetic type of each individual at a given locus is characterized by the allelic types of its genes at this locus, *i.e.*, its single-locus genotype. Within the population, the form of association of the alleles in the gene-pool into single-locus genotypes determines the distribution of single-locus genotypes. Random association yields Hardy-Weinberg proportions.

7.1.3 Multilocus genotypes

When considering more than one gene locus simultaneously, the genetic type of an individual is characterized by the list of its single-locus genotypes at each of these loci, *i.e.*, its multilocus genotype. Within the population, the form of association of the single-locus genotypes of the individuals into multilocus genotypes determines the distribution of multilocus genotypes. Random association appears as gametic equilibrium.

7.2 Measures considering elementary genic differences

At each level of the three levels of genetic integration, *DifferInt* calculates the following measures. These measures are based not only on relative frequencies of the genetic types of the individuals in the populations but also on the degree to which the genetic types themselves differ from each other.

7.2.1 Elementary genic difference *d* between genetic types

The genic difference between two genetic types at the same level of integration is basically determined by the number of their individual genes that differ in allelic type. If the numbers of copies of the *i*-th allele at the *l*-th gene locus are denoted by $n_{i;l}$ and $m_{i;l}$, respectively, then the two genetic types differ by $\sum_{i,l} |n_{i;l} - m_{i;l}|$ gene-type copies. This sum is maximal, equaling two times the total number K of individual genes represented in each object, if the objects share no gene-types (and thus differ completely). Since $\sum_{i,l} n_{i;l} =$ $\sum_{i,l} m_{i;l} = K$ holds, division of $\sum_{i,l} |n_{i;l} - m_{i;l}|$ by 2K yields a measure of genic difference that is bounded between zero and one. This measure of elementary genic difference is applicable to all levels of integration.

7.2.2 Pairwise genetic distance Δ between populations

The pairwise genetic distance

$$\Delta(\mathcal{P}, \mathcal{Q})$$

equals the minimum degree to which the frequency distribution of the genetic types within one of the populations, say \mathcal{P} , must be transformed in order to make it match the distribution in the other, say \mathcal{Q} , where the degree of transformation is based on the elementary genic differences between genetic types (Gregorius *et al.*, 2003; Gillet *et al.*, 2004).

A transformation s consists of a set of frequency shifts within population \mathcal{P} from types that are more frequent in population \mathcal{P} than in \mathcal{Q} to types that are less frequent in \mathcal{P} than in \mathcal{Q} for the purpose of making population \mathcal{P} match population \mathcal{Q} . If the frequency p_a of type a in \mathcal{P} exceeds the frequency q_a of this type in \mathcal{Q} , then the excess $p_a - q_a$ must be shifted to types deficient in \mathcal{P} , such that $\sum_b s(a,b) = p_a - q_a = p_a - \min\{p_a,q_a\}$. The shift process is continued for all types with a frequency excess in \mathcal{P} until the frequencies of all types in \mathcal{P} match those in \mathcal{Q} . Denoting s as the transformation, *i.e.*, the totality of frequency shifts, and weighting each shift s(a,b) by the elemtary genic difference d(a,b) between the respective genetic types yields

$$\Delta(s) = \sum_{a,b} s(a,b) \cdot d(a,b)$$

 $\Delta(s)$ can be considered as the "cost" of transformation s.

It is easy to see that at least one such transformation exists. Since there may be more than one, Δ is taken to be the minimum value of $\Delta(S)$ over all permissible transformations S, *i.e.*,

$$\Delta(\mathcal{P}, \mathcal{Q}) = \min_{S} \Delta(S)$$

Gregorius *et al.* (2003) and Gillet *et al.* (2004) show that finding a shift transformation *s* that minimizes $\Delta(s)$ is equivalent to solving the cost-minimizing *Transportation Problem* (Hitchcock, 1941) of operations research. Linear programming methods for its solution are implemented in the computer program *DeltaS* (http://www.uni-goettingen.de/de/95605.html).

See App. A for an explanation of special relationships between values of the genetic difference Δ when calculated at different levels of genetic integration (gene-pool, mean single-locus genotypes, and multilocus genotypes).

DifferInt outputs lower triangular matrices containing the pairwise distances Δ for all pairs of populations. These distance matrices can be input into

other programs that make dendograms or choropleths, for example.

7.2.3 Complementary subpopulation differentiation Δ_{SD}

The complementary subpopulation differentiation

$$\Delta_{SD} = \sum_{j=1}^{n} c_j \cdot \Delta_{SD}(j)$$

(Gillet & Gregorius, 2008) equals the weighted arithmetic mean of the genetic distances

$$\Delta_{SD}(j) = \Delta(\mathcal{P}(j), P^c(j))$$

of each subpopulation $\mathcal{P}(j) = (p_1(j), \ldots, p_n(j))$ to its complement population $\mathcal{P}^c(j) = (p_1^c(j), \ldots, p_n^c(j))$ that is formed by pooling all other populations, where $p_i^c(j) = \sum_{k \neq j} p_i(k)/(1 - c_j)$ and the weight c_j is the relative size of subpopulation $\mathcal{P}(j)$. In DifferInt, subpopulation sizes are disregarded, yielding

$$c_j \equiv 1/n$$
 and $p_i^c(j) = \sum_{k \neq j} p_i(k) \cdot n/(n-1)$

The name of this measure indicates that it was conceived for the measurement of differentiation among subpopulations belonging to a larger population, but it is applicable to any set of populations. Δ_{SD} ranges between 0 and 1. Δ_{SD} equals 1 if all populations are genetically disjoint, *i.e.*, no two populations share the same genetic types. Δ_{SD} equals 0 if all populations show identical frequency distributions for the genetic types.

7.2.4 Dispersive subpopulation differentiation Δ

The dispersive subpopulation differentiation

$$\bar{\Delta} = \sum_{j=1}^{n} c_j \cdot \bar{\Delta}(j)$$

equals the weighted arithmetic mean of the mean genetic distance Δ of each population $\mathcal{P}(j)$ to each of the others, *i.e.*,

$$\bar{\Delta}(j) = \frac{1}{1 - c_j} \cdot \sum_{k \neq j} c_k \cdot \Delta(\mathcal{P}(j), \mathcal{P}(k))$$

of each subpopulation $\mathcal{P}(j) = (p_1(j), \ldots, p_n(j))$ to each of the others and the weight c_j is the relative size of subpopulation $\mathcal{P}(j)$. In *DifferInt*, subpopulation sizes are disregarded, yielding

$$c_j \equiv 1/n$$
 and $p_i^c(j) = \sum_{k \neq j} p_i(k) \cdot n/(n-1)$

The name of this measure indicates that it is an average. $\overline{\Delta}$ ranges between 0 and 1. $\overline{\Delta}$ equals 1 if all populations are genetically disjoint, *i.e.*, no two populations share the same genetic types. $\overline{\Delta}$ equals 0 if all populations show identical frequency distributions for the genetic types.

7.3 Measures neglecting elementary genic differences

At each level of the three levels of genetic integration, *DifferInt* also calculates the following measures. These are equivalent to the measures in the previous sections with the exception that the differences between genetic types are neglected. By replacing the elementary genic difference between genetic types by the discrete difference, the measures given below are based only on relative frequencies of the genetic types of the individuals in the populations.

7.3.1 Discrete difference between genetic types

The discrete difference between two genetic types at the same level of genetic integration equals 0 if the types are identical over all alleles at all of the loci under consideration and 1 if the types differ by at least one allele at one locus.

7.3.2 Pairwise genetic distance d_0 between two populations

The pairwise genetic distance

$$d_0(\mathcal{P}, \mathcal{Q}) = \frac{1}{2} \sum_{i=1}^n |p_i - q_i|$$

(Gregorius, 1974) between populations \mathcal{P} of frequency distribution $\mathcal{P} = (p_1, \ldots, p_n)$ and \mathcal{Q} of frequency distribution $\mathcal{Q} = (q_1, \ldots, q_n)$ equals the minimum degree to which the frequency distribution of the genetic types within one of the populations must be transformed in order to make it match the distribution of types in the other population (Gregorius *et al.*, 2003; Gillet *et al.*, 2004; Gregorius, 1987, 1984).

 Δ equals d_0 if the "cost" of transformation for Δ is taken to be the discrete difference. Values of d_0 range between 0 and 1. d_0 equals 1 if the two populations are genetically disjoint, *i.e.*, do not share any genetic type. d_0 equals 0 if $\mathcal{P} = Q$.

DifferInt outputs lower triangular matrices containing the pairwise distances d_0 for all pairs of populations. These distance matrices can be input into other programs that make dendograms or choropleths, for example.

7.3.3 Complementary subpopulation differentiation δ_{SD}

The complementary subpopulation differentiation

$$\delta_{SD} = \sum_{j=1}^{n} c_j \cdot D(j)$$

(Gregorius, 1996; Gregorius & Roberds, 1986) equals the weighted arithmetic mean of the genetic distances

$$\delta_{SD}(j) = D_j = d_0(\mathcal{P}(j), \bar{P}(j)) = \frac{1}{2} \sum_{i=1}^n |p_i(j) - \bar{p}_i(j)|$$

of each subpopulation $\mathcal{P}(j) = (p_1(j), \dots, p_n(j))$ to its complement population $\bar{P}(j) = (\bar{p}_1(j), \dots, \bar{p}_n(j))$ that is formed by pooling all other populations,

where $\bar{p}_i(j) = \sum_{k \neq j} p_i(k)/(1 - c_j)$ and the weight c_j is the relative size of subpopulation $\mathcal{P}(j)$. In *DifferInt*, subpopulation sizes are disregarded, yielding

$$c_j \equiv 1/n$$
 and $\bar{p}_i(j) = \sum_{k \neq j} p_i(k) \cdot n/(n-1)$

 Δ_{SD} equals δ_{SD} if the "cost" of transformation for the pairwise difference Δ is taken to be the discrete difference. Values of δ_{SD} range between 0 and 1. δ_{SD} equals 1 if all populations are genetically disjoint, *i.e.*, no two populations share the same genetic types. δ_{SD} equals 0 if all populations show identical frequency distributions for the genetic types.

7.3.4 Dispersive subpopulation differentiation $\bar{\delta}$

The dispersive subpopulation differentiation

$$\bar{\delta} = \sum_{j=1}^{n} c_j \cdot \bar{\delta}(j)$$

equals the weighted arithmetic mean of the mean genetic distance d_0 of each population $\mathcal{P}(j)$ to each of the others, *i.e.*,

$$\bar{\delta}(j) = \frac{1}{1 - c_j} \cdot \sum_{k \neq j} c_k \cdot d_0(\mathcal{P}(j), \mathcal{P}(k))$$

of each subpopulation $\mathcal{P}(j) = (p_1(j), \ldots, p_n(j))$ to each of the others and the weight c_j is the relative size of subpopulation $\mathcal{P}(j)$. In this version of *DifferInt*, subpopulation sizes are considered equal. $\bar{\delta}$ ranges between 0 and 1. $\bar{\delta}$ equals 1 if all populations are genetically disjoint, *i.e.*, no two populations share the same genetic types. $\bar{\delta}$ equals 0 if all populations show identical frequency distributions for the genetic types.

7.4 Covariation of differentiation between integration levels

The degree of correspondence between differentiation indices from two levels of integration can be determined by a measure of covariation. As was pointed out in Gregorius et al. (2007), a suitable measure of covariation is

$$C = \frac{\sum_{i < k} (X_i - X_k) (Y_i - Y_k)}{\sum_{i < k} |(X_i - X_k) (Y_i - Y_k)|}$$

where the variables X_j and Y_j refer to the contribution of population $\mathcal{P}(j)$ at two different levels of integration and X_k and Y_k to the respective contributions of a different population k. In the case of complementary differentiation Δ_{SD} and δ_{SD} , X_j and Y_j refer to $\Delta_{SD}(j)$ and $\delta_{SD}(j)$, respectively, at two levels of integration. In the case of dispersive differentiation $\overline{\Delta}$ and $\overline{\delta}$, X_j and Y_j refer to $\overline{\Delta}(j)$ and $\overline{\delta}(j)$, respectively, at two levels. C varies between -1 and +1 such that C = 1 for strictly positive and C = -1 for strictly negative covariation. It is undefined in the practically irrelevant case where a non-zero difference for one variable implies equality for the other.

7.5 Statistical analysis

The significance of the observed measures of distance and differentiation at each level of integration is estimated by two kinds of permutation analysis.

7.5.1 Testing random association of genes to genotypes within populations

As stated by Gillet & Gregorius (2008), "Any increase of genetic differentiation among populations at higher levels of genetic integration is due to forces of association of genes that differ among populations. It is thus of basic interest to know whether the differentiation observed at a level of integration can be explained by random combination of genes (e.g. into diploid genotypes or haplotypes) or whether directed forces of combination must be assumed. This requires an analysis that is conditional on the gene-pool of each population, the number of populations, and the population sizes. The effects of chance can be assessed by permuting the genes within each population, such that all homologous and non-homologous combinations of genes (alleles) into (haploid, diploid or polyploid) genotypes have equal probability."

For each permutation, at each gene locus the genes present within each population are randomly reassigned to the individuals, creating new distributions of single- and multilocus genotypes within populations and, accordingly, new realizations of the associations of the genetic types at one level of genetic integration into the genetic types at the next higher level (*i.e.*, gene-pool to single-locus genotypes, single-locus genotypes to multilocus genotypes).

The values of all measures at each integration level are calculated for each permutation. After all permutations have been completed, the P-value for each measure is determined as the proportion of all permutations that yielded values greater than or equal to the observed value. For interpretation of the results, both very small P-values (0.05) and very large P-values (0.95) are of interest. *DifferInt* prints the following results from the permutations for each measure (see Tab.2):

- observed differentiation
- minimum value of differentiation over all permutations
- lower and upper bounds of the confidence interval as the central X% of the values from all permutations (where "X" is the chosen "Size of confidence intervals as percentage of permutation")
- maximum value of differentiation over all permutations
- P-value for the observation

It may occur that the hypothesis is rejected at one level of integration but not another. This can provide clues to the cause of the non-random association.

7.5.2 Testing random assignment of genetic types to populations

Populations are differentiated when genetic types are unevenly distributed over populations. To test whether genetic types to populations could have been randomly assigned to the populations, the (multilocus) genotypes of all individuals in the metapopulation are randomly permuted among the populations, resulting in new assignments of genetic types to populations. The total number of individuals with each genetic type at each level of integration remains the same.

For each permutation, *DifferInt* calculates the four differentiation measures at each integration level. After completion of all permutations, the analogous results as in Sec. 7.5.1 are listed in the output (see Tab.2). Rejection of the hypothesis at one level of integration but not another can provide clues to the cause of the non-random assignment.

8 Output files

Besides appearing in a window, results are printed to the following files, where the prefix "aaa" is that specified during program initiation:

- * aaa.summary.txt
 - SUMMARY OF OBSERVATIONS AND ALL PERMUTATIONS
 - DIFFERENTIATION at levels of genetic integration GenePool, MnSLGeno, and MultGeno
 - COVARIATION OF DIFFERENTIATION between levels of integration
 - > Observed values
 - > Permutation analysis: Minimum, Confidence Interval, Maximum, P-Value using differentiation measures that consider elementary genic differences
 - > Complementary population differentiation Δ_{SD}
 - > Dispersive differentiation $\bar{\Delta}$
 - > Complementary population differentiation δ_{SD}
 - > Dispersive differentiation $\bar{\delta}$
- * aaa.alleles.txt, aaa.singlgeno.txt, and aaa.multigeno.txt More detailed output for the each of the three levels of integration
 - > Frequency distributions of the genetic types in the populations
 - > Matrices of elementary genic differences between genetic types
 - > Optimal shift matrices of the distance measure Δ for all comparisons between two populations and between each population and its complement population at all levels of genetic integration

* aaa.excluded.txt

List of (multilocus) genotypes that are excluded from the analysis due to missing data

- * aaa.dOmatrix.txt and aaa.Deltamatrix.txt List of distance matrices for the different levels of genetic integration
- * aaa.bigdeltasd.schnecken.fig, aaa.bigdeltabar.schnecken.fig, aaa.deltasd.schnecken.fig, aaa.deltabar.schnecken.fig

Input files for plot software xfig (Linux) or winfig (Windows). The fig-files are constructed from the files of the same names ending in .in. If the answer to Make snail graphics is "yes", the .fig-files are plotted on the screen and can be exported in various formats. See Figs. 4-7.











Figure 6: Snails for the complementary differentiation measure δ_{SD} that neglects elementary genic differences. Input file exal.txt.



elementary genic differences. Input file exa1.txt. Figure 7: Snails for the dispersive differentiation measure $\overline{\delta}$ that neglects

9 Compilation

9.1 gfortran compiler

DifferInt is available as executables for Windows and Linux.

9.2 Windows

Compiled for Windows-XP (32-bit) using the GNU Fortran 95 compiler gfortran (http://gcc.gnu.org/fortran), version 4.8.0 20120518 (experimental) [trunk revision 187663], built for MinGW.

9.3 Linux

Compiled for Linux (32-bit) using the Fortran 95 compiler gfortran (http://gcc.gnu.org/fortran), version 4.7.1 20120723 [gcc-4_7-branch revision 189773], built on opensuse 12.2.

9.4 Implemented software

DifferInt implements subroutines from the following software:

9.4.1 *RELAX-IV*

DifferInt incorporates open-source code from the program RELAX-IV Code for Solving the Minimum Cost Network Flow Problem of Bertsekas & Tseng (1994) that solves the Hitchcock-Koopmans Transportation Problem known from the field of logistics by linear programming methods. This method is used to calculate the pairwise distance Δ between two populations by minimizing the extent to which the genetic types of individuals in one of the populations must be altered in order to make this population match the composition of genetic types in the other.

9.4.2 init_random_seed()

DifferInt applies the pseudorandom number generator RANDOM_NUMBER of open-source GNU Fortran (http://gcc.gnu.org/fortran). The random seed is initialized based on the computer system's time and thus differs for each run. The applied subroutine init_random_seed() is found on pp. 185ff of the manual by The gfortran team (1999-2013).

9.4.3 DISLIN

Widgets for the input and output windows are generated by routines of the *DISLIN* Scientific Plotting Software by H. Michels (http://www.dislin.de).

9.4.4 WinFIG and xfig

Snail diagrams are plotted using vector graphics software. For Windows, the software package *WinFIG* (http://www.winfig.com) must be installed on the computer. For Linux, the packages *xfig* and *transfig* (http://www.xfig. org) must be installed.

10 Copyright and terms of use

DifferInt - Compositional differentiation among populations at three levels of genetic integration.

(C) 2008-2013 Elizabeth M. Gillet (Email: egillet@gwdg.de) excepting code from RELAX-IV of Bertsekas & Tseng (1994) and init_random_seed() of The gfortran team (1999-2013).

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Send comments, suggestions, and bug reports to differint@uni-goettingen.de.

Acknowledgements: This work was funded by grant Zi 662/5-2 of the Deutsche Forschungsgemeinschaft.

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A Comparing genetic distance Δ at different levels of genetic integration

For a given set of gene loci, the following always holds for the genetic distances $\Delta(\mathcal{P}, \mathcal{Q})$ between any two populations \mathcal{P} and \mathcal{Q} at the different levels of genetic integration:

- * The mean over loci of the distances $\Delta(\mathcal{P}, \mathcal{Q})$ between the allelic frequency distributions (gene-pool) cannot be greater than the mean over loci of the distances $\Delta(\mathcal{P}, \mathcal{Q})$ between the frequency distributions of the single-locus genotypes.
- * The mean over loci of the distances $\Delta(\mathcal{P}, \mathcal{Q})$ between the frequency distributions of the single-locus genotypes cannot be greater than the distance $\Delta(\mathcal{P}, \mathcal{Q})$ between the frequency distributions of the multilocus genotypes.

In some special cases, $\Delta(\mathcal{P}, \mathcal{Q})$ does not increase from a lower to a higher level of genetic integration:

Case 1: For a given gene locus, let the genotypes of the individuals in both population \mathcal{P} and population \mathcal{Q} show Hardy-Weinberg proportions. The distributions of the alleles may differ between the populations. Then the following measures are equal:

 $\Delta(\mathcal{P}, \mathcal{Q})$ at the integration level of the single-locus genotypes = $\Delta(\mathcal{P}, \mathcal{Q})$ at the integration level of the gene-pool (alleles) = $d_0(\mathcal{P}, \mathcal{Q})$ at the level of the gene-pool.

The same equality holds if \mathcal{P} and \mathcal{Q} both show inbreeding structures with the same value of the inbreeding coefficient F (Hardy-Weinberg proportions correspond to inbreeding structures with F = 0.)

Case 2: For a given set of gene loci, let the multilocus genotypes of the individuals in population \mathcal{P} and in population \mathcal{Q} show random association of the single-locus genotypes among gene loci. The distributions of the multilocus genotypes may differ between the populations. Then the following measures are equal: $\Delta(\mathcal{P},\mathcal{Q})$ at the integration level of the multilocus genotypes = $\Delta(\mathcal{P},\mathcal{Q})$ at the integration level of the mean single-locus genotypes

Combination of Cases 1 and 2: For a given set of gene loci, let both population \mathcal{P} and population \mathcal{Q} show Hardy-Weinberg proportions (or equal inbreeding coefficients F) at all loci and let the single-locus genotypes be randomly associated in the multilocus genotypes of the individuals within each population. Then the following measures are equal:

 $\Delta(\mathcal{P}, \mathcal{Q})$ at the integration level of the multilocus genotypes = $\Delta(\mathcal{P}, \mathcal{Q})$ at the integration level of the mean single locus genotypes = $d_0(\mathcal{P}, \mathcal{Q})$ at the integration level of the gene-pool.

B Example of an output file

The following pages (Tab. 2) show the output file aaa.summary.txt that resulted from the example input file exal.txt. The prefix aaa of the file name is user-specified, the size of confidence intervals equals 95% of the permutation values, and number of permutations equals 1000. Because initiation of the random number generator is based on the system's time, the results of permutation analysis change from run to run, but they should stabilize as the number of permutations is increased. Table 2: Output file aaa.summary.txt for input file exa1.txt and prefix aaa. Continued on next pages.

: DifferInt (May 2013) compiled by GNU Fortran (SUSE Linux) 4.3.2 Program Input file : exal.txt Begin : 16.05.2013 at 10:32:46 OBSERVED_DIFFERENTIATION_OVER_THREE_LEVELS_OF_INTEGRATION FOR LOCI 1 2 Sample_size = No._individuals_with_no_missing_alleles Population_weight = 1/No.populations 2 Pop 1 3 22 SmplSiz 10 18 Weight 0.333 0.333 0.333 GENE_POOL Complementarv Dispersive Pairwise differentiation differentiation distances Popj Delta_SD(j) Delta-bar(j) Delta =delta SD(i) =delta-bar(i) =d03 2 1 0.2095 1 0.1564 0.0000 0.0705 0.3485 2 0.2424 0.2436 0.0705 0.0000 0.4167 3 0.3826 0.3826 0.3485 0.4167 0.0000 Delta-bar Delta_SD =delta_SD =delta-bar 0.2785 0.2605 MEAN SINGLE LOCUS GENOTYPES - CONSIDERING ELEMENTARY GENIC DIFFERENCES Complementary Dispersive Pairwise differentiation differentiation distances Popj Delta_SD(j) Delta-bar(j) Delta 2 3 1 0.0000 0.0932 1 0.1977 0.2455 0.3977 0.2549 2 0.2424 0.0932 0.0000 0.4167 3 0.4053 0.4072 0.3977 0.4167 0.0000 Delta SD Delta-bar 0.2818 0.3025

MEAN_SINGLE_LOCUS_GENOTYPES - NEGLECTING_ELEMENTARY_GENIC_DIFFERENCES Complementary Dispersive Pairwise differentiation differentiation distances Popi delta_SD(j) delta-bar(j) d0 1 2 3 0.2742 0.3202 0.0000 0.1227 0.5177 1 2 0.3232 0.3391 0.1227 0.0000 0.5556 3 0.5328 0.5366 0.5556 0.5177 0.0000 delta SD delta-bar 0.3987 0.3768 MULTILOCUS GENOTYPES - CONSIDERING ELEMENTARY GENIC DIFFERENCES Dispersive Complementary Pairwise differentiation differentiation distances Popj Delta_SD(j) Delta-bar(j) Delta 1 2 3 0.2396 0.2659 0.0000 0.1227 0.4091 1 0.1227 0.0000 2 0.2427 0.2711 0.4194 3 0.4092 0.4143 0.4091 0.4194 0.0000 Delta SD Delta-bar $0.297\bar{2}$ 0.3171 MULTILOCUS GENOTYPES - NEGLECTING ELEMENTARY GENIC DIFFERENCES Complementary Dispersive Pairwise differentiation differentiation distances Popj delta_SD(j) delta-bar(j) d0 3 2 1 0.5273 1 0.5182 0.0000 0.2364 0.8182 2 0.4354 0.5126 0.2364 0.0000 0.7889 3 0.7934 0.8035 0.8182 0.7889 0.0000 delta SD delta-bar 0.5823 0.6145

=======================================			===			=======	=======
P_E_F PERMUTATION_OF_ALLEI	R_M_U_T_A_3 LES_OVER_IN	C_I_O_N NDIVIDUAL	A_ _s	N_A_L_Y_S_ W_I_T_H_I_	I_S NPOPULATI	ONS	
1000 PERMUTATIONS_OF_ALLELES_OV	/ER_INDIVII	DUALSW_	I_T	_H_I_NPO	PULATIONS		
AT_LEVELS_OF_GENETIC_INTEGRATION GenePool							
	MnSLGer MultGer	1o=Mean_S 1o=Multil	ocu	s Genotype	enotypes s		
DIFFERENTIATION	Ubserved values	Min	0.9	Permut 50-Confide	ation_analy nceInterval	sis Max	P-Value
Considering_elementary_genic_di	ifferences	:					
Complementary-Differentiation-I	Delta_SD						
GenePool	$0.\overline{2}605$			not	_affected .		
MnSLGeno	0.2818	0.2605	[0.2626 ,	0.3083]	0.3324	0.3000
MultGeno	0.2972	0.2658	Ē	0.2776 ,	0.3275]	0.3479	0.5430
Dispersive-Differentiation-Delt	ta-bar			-			
GenePool	0.2785			not	_affected .		
MnSLGeno	0.3025	0.2805	[0.2830 ,	0.3396]	0.3626	0.5010
MultGeno	0.3171	0.2865	Γ	0.3002 ,	0.3524]	0.3742	0.6560
Neglecting_elementary_genic_dif	fferences:						
Complementary-Differentiation-c	delta_SD						
GenePool	0.2605			not	_affected .		
MnSLGeno	0.3768	0.3710	[0.3932 ,	0.5177]	0.5729	0.9950
MultGeno	0.5823	0.5658	[0.6195 ,	0.8116]	0.9212	0.9980
Dispersive-Differentiation-delt	ta-bar						
GenePool	0.2785		· <u>-</u> ·	not	_affected .	• • • • • • • • •	••••••••
MnSLGeno	0.3987	0.3867	Ľ	0.4264 ,	0.5746]	0.6338	0.9970
MultGeno	0.6145	0.6101	L	0.6653 ,	0.8539]	0.9394	0.9990
COVARIATION OF DIFFERENTIATION	Observed			Permut	ation analy	 sis	
···· · _ - _ · ·	values	Min	0.9	50-Confide	nceInterval	Max	P-Value
Considering_elementary_genic_di	ifferences	:					
Cov(Delta_SD(j))							
GenePool-MnSLGeno	1.0000	0.7995	Γ	1.0000 .	1.0000]	1.0000	0.0000
GenePool-MultGeno	1.0000	0.8387	Ī	1.0000 .	1.0000 1	1.0000	0.0000
MnSLGeno-MultGeno	1.0000	0.9912	Ī	1.0000	1.0000 1	1.0000	0.0000
Cov(Delta-bar(j))			-	,			
GenePool-MnSLGeno	1.0000	0.9479	Г	0.9969	1.0000]	1.0000	0.000
GenePool-MultGeno	1,0000	0.9397	ř	0.9969	1,0000 1	1.0000	0.0000
MnSLGeno-MultGeno	1,0000	0.9904	ř	1.0000	1,0000 1	1.0000	0.0000
Neglecting elementary genic dif	fferences:		-	,]		
Cov(delta SD(i))							
ConoPool MnSI Cono	1 0000	0 7404	Г	0 0800	1 0000]	1 0000	0 0000
ConcDool Mul+Conc	1.0000	0.7404 0.7107	L F	0.3030,	1 0000]	1 0000	0.0000
Mngi Cono MultCono	0.0009	0.1191	L F	0.3000,	1 0000]	1 0000	0.9010
Cov(dolta bar(i))	0.9400	0.0102	L	0.3430 ,	T.0000]	1.0000	0.9140
ConcDeal Madi Cara	1 0000	0 0165	г	0 0047	1 0000 7	1 0000	0 0000
ConcDecl MultCere	1.0000	0.9100	L	0.9941, 0.0711	1 0000]	1 0000	0.0000
MnSI Cono MultCono	0.9007	0.0930	L L	0.9741,	1 0000 1	1 0000	0.3340
LIDERGEIIO-LITE CAGILO	0.3300	0.1200	L	0.0010,	T.00000]	1.0000	0.0120

P_E_R_M_U_T_A_T_I_O_NA_N_A_L_Y_S_I_S									
PERMUTATION_OF_ALL_INDIVIDUAL_GENOTYPESA_M_O_N_GTHE_POPULATIONS									
1000 PERMUTATIONS_OF_ALL_INDIVIDUAL_GENOTYPESA_M_O_N_GPOPULATIONS FOR ALL 2 LOCT									
AT_LEVELS_OF_GENETIC_INTEGRATI	AT_LEVELS_OF_GENETIC_INTEGRATION GenePool								
MnSLGeno=Mean_Single_Locus_Genotypes MultGeno=Multilocus_Genotypes									
DIFFERENTIATION	values	Min O	.950-Confide	nceInterval	Max	P-Value			
Considering_elementary_genic_d	ifferences	:							
Complementary-Differentiation-	Delta_SD	0 0000	F 0 0000	0 0500]	0 0040	0.0450			
GenePool MnSI Cono	0.2605	0.0662	$\begin{bmatrix} 0.0998 \\ 0.1292 \end{bmatrix}$	0.2528]	0.3018	0.0150			
MultGeno	0.2010	0.1011	$\begin{bmatrix} 0.1305 \\ 0.1675 \end{bmatrix}$	0.2796]	0.3239	0.0230			
Dispersive-Differentiation-Del	ta-bar	0.1010	2 0.1010 ,	0.2000]	0.0101	0.0200			
GenePool	0.2785	0.0801	[0.1137 ,	0.2862]	0.3391	0.0420			
MnSLGeno	0.3025	0.1198	L 0.1598 ,	0.3198	0.3665	0.0510			
MultGeno Noglocting olomontary gonic di	U.3171	0.1545	[0.1942 ,	0.3378]	0.3774	0.0780			
Complementary-Differentiation-	delta_SD								
GenePool	0.2605	0.0662	[0.0998 ,	0.2528]	0.3018	0.0150			
MnSLGeno	0.3768	0.1535	[0.2033 ,	0.3817]	0.4786	0.0310			
MultGeno 0.5823 0.3386 [0.4125 , 0.6291] 0.7093 0.1340									
Ulspersive-Uliferentiation-delta-bar									
MnSL.Geno	0.3987	0.1815	$\begin{bmatrix} 0.1137 \\ 0.2337 \end{bmatrix}$	0.4328]	0.5153	0.1050			
MultGeno	0.6145	0.3859	[0.4714 ,	0.6956]	0.7828	0.3050			
Output files:	==========	===========	===========		=======	======			
aaa.summary.txt									
aaa.alleles.txt									
aaa.singiegeno.txt									
aaa.dOmatrix.txt : pair	wise dista	nces d0 be	etween pops a	t all level	S				
aaa.Deltamatrix.txt : pairwise distances Delta between pops at all levels									
To_plot_snails,_enter_the_following_commands_in_a_terminal_window:									
xfig -allownegcoords aaa.bi	gdeltasd.s	chnecken.f	ig						
xiig -allownegcoords aaa.de	odeltabar	ecken.11g schnecker	fiσ						
xfig -allownegcoords aaa.deltabar.schnecken.fig									
End: 16.05.2013 at 10:32:58									